



AFNI



# Didactics and Demonstrations

Hands-On Session: Regression Analysis

[File: afni05\\_regression.pdf](#)

# Overview

## • What we have learned so far

- Use data viewer 'afni' interactively
- Model HRF with a **fixed-shape** basis function (*e.g.*, Gamma variate)
  - Assume the brain responds with the **same shape**
    - Across **subjects** and any activated **regions**
  - Differ in **magnitude**:  $\beta$  (and its significance) is focus

## • What we will do in this hands-on session

- Data pre-processing overview for time series regression analysis
- Basic concepts
  - Regressors, design matrix, and confounding effects
  - Statistical significance testing in regression analysis
- Navigation with GUI '**afni**'
  - Spot check for the original data
  - Statistic thresholding with data viewer '**afni**' (**two-sided** vs. **one-sided** with  $t$ )
  - Model performance (visual check of curve fitting and test via full  $F$  or  $R^2$ )

# FMRI Regression Analysis

- Voxel-wise regression model:  $y = X\beta + \varepsilon$ 
  - $y$ : signal (time series) at a voxel - **different** across voxels
  - $X$ : explanatory (independent) variables (regressors) - **same** across voxels
  - $\beta$ : regression coefficients (response strength) - **different** across voxels
  - $\varepsilon$ : residuals (anything we can't account for) - **different** across voxels
- Regressors in design matrix  $X = [x_1, x_2, \dots, x_k]$ 
  - Regressors of interest: hemodynamic responses (HDR)
  - Regressors of no interest: drift effect (polynomials), head motion, *etc.*
- Association between stimulus and BOLD signal: HDR/HRF
  - Fixed shape regardless of subjects, brain regions, stimuli: regression
  - No assumption about the HDR shape: deconvolution + regression
  - Middle ground: regression
- Residuals
  - White noise: OLS - 3dDeconvolve
  - Serially correlated: ARMA(1,1)+REML - 3dREMLfit

# A Case Study

- ◆ **Speech Perception Task:** Subjects were presented with audiovisual speech presented in a predominantly auditory or predominantly visual modality.
- ◆ A digital video system was used to capture auditory and visual speech from a female speaker.
- ◆ 2 types of stimulus conditions:



(1) **Auditory-Reliable**

Example: Subjects can clearly *hear* the word “cat,” but the video of a woman mouthing the word is degraded.

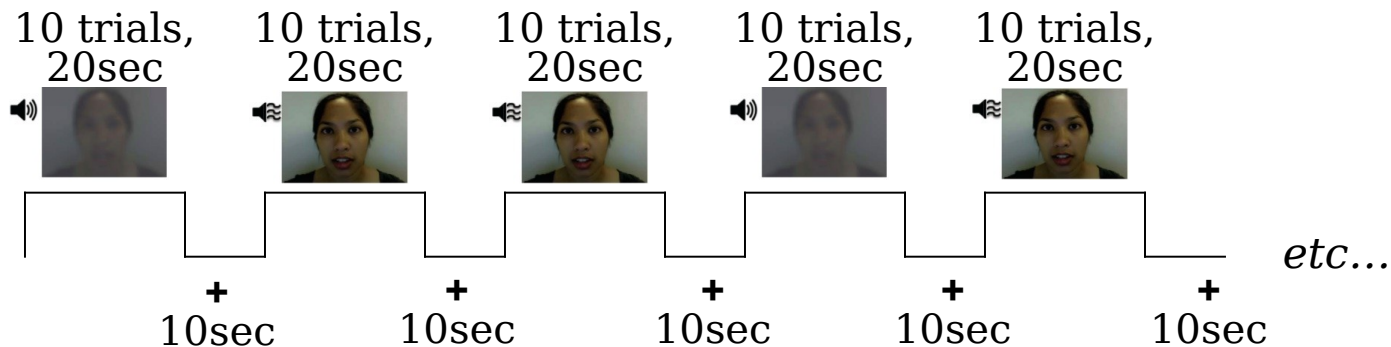


(2) **Visual-Reliable**

Example: Subjects can clearly *see* the video of a woman mouthing the word “cat,” but the audio of the word is degraded.

# Experiment Design

- ◆ 3 runs in a scanning session
- ◆ Each run consisted of randomized 10 blocks:
  - 5 blocks contained Auditory-Reliable (*Arel*) stimuli, and
  - 5 blocks contained Visual-Reliable (*Vrel*) stimuli
- ◆ Each block contained 10 trials of *Arel* OR *Vrel* stimuli
  - Each block lasted for 20s (1s for stimulus presentation, followed by a 1s inter-stimulus interval)
- ◆ Each baseline block consisted of a 10s fixation point

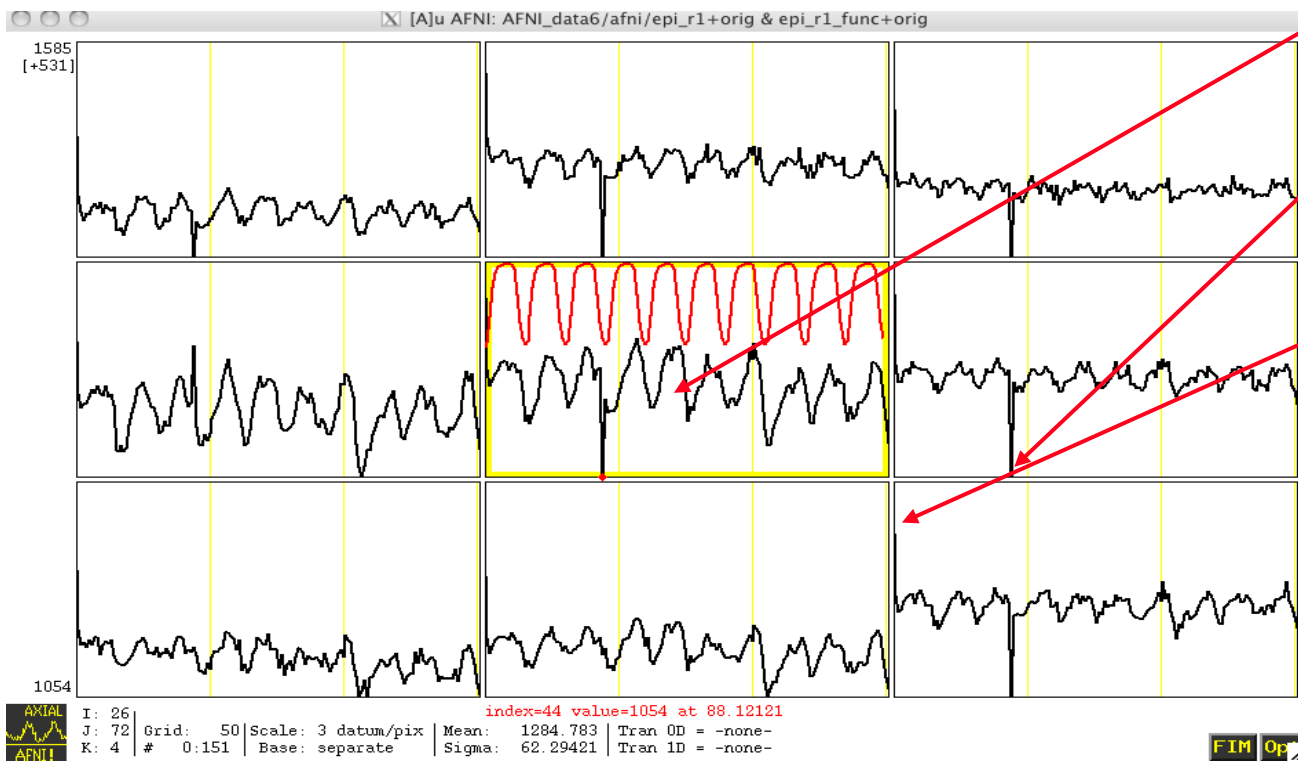


# Data Collected

- ◆ 2 anatomical datasets for each subject from a 3T
  - 175 sagittal slices
  - voxel dimensions =  $1.0 \times 0.938 \times 0.938 \text{ mm}^3$
- ◆ 3 time series (EPI) datasets for each subject
  - 33 axial slices  $\times$  152 volumes (TRs) per run
  - TR = 2s; voxel dimensions =  $2.75 \times 2.75 \times 3.0 \text{ mm}^3$
- ◆ Sample size,  $\underline{n} = 10$  (all right-handed subjects)
  - 10 is far too few for most modern experiments

# Data Quality Check

- To look at the data: type **cd AFNI\_data6/afni**, then **afni**
- **Switch Underlay** to dataset **epi\_r1**
  - Then **Axial Image** and **Graph**
  - **FIM→Pick Ideal** ; then click **afni/epi\_r1\_ideal.1D** ; then **Set**
  - Right-click in image, **Jump to (ijk)**, then **26 72 4**, then **Set**



- Data clearly has activity in sync with reference
  - 20s blocks
- Data also has a big spike at 89s
  - Head motion
- Spike at  $t = 0$
- Some tricks with keyboard
  - **oa**: automatic scaling
  - **ov**: video mode
  - **om/M**: voxel matrix sizing on Graph window

# Preparing Data for Analysis

- Following preparatory steps are common (e.g., `afni_proc.py`):
  - Outliers: `3dToutcount`, `3DDespike`
  - Temporal alignment or slice timing correction (sequential/interleaved): `3dTshift`
  - EPI Image/volume registration (head motion correction): `3dvolreg`
  - EPI to anatomy registration: `align_epi_anat.py`
  - Spatial normalization (standard space conversion): `@auto_tlrc`, `auto_warp.py`
  - Blurring/smoothing: `3dmerge`, `3dBlurToFWHM`, `3dBlurInMask`
  - Masking: `3dAutomask`
  - Temporal mean scaling: `3dTstat` and `3dcalc`
- Not all steps are necessary or desirable in any given case



# Regression Analysis

- Regression model:  $y = X\beta + \varepsilon$
- Run script by typing **tcsh rall\_regress** (takes a few minutes)

```

3dDeconvolve -input rall_vr+orig -polort 1 \
  -concat '1D: 0 150 300' \
  -num_stimts 8 \
  -stim_times 1 stim_AV1_vis.txt 'BLOCK(20,1)' -stim_label 1 Vrel \
  -stim_times 2 stim_AV2_aud.txt 'BLOCK(20,1)' -stim_label 2 Arel \
  -stim_file 3 motion.1D'[0]' -stim_base 3 -stim_label 3 roll \
  -stim_file 4 motion.1D'[1]' -stim_base 4 -stim_label 4 pitch \
  -stim_file 5 motion.1D'[2]' -stim_base 5 -stim_label 5 yaw \
  -stim_file 6 motion.1D'[3]' -stim_base 6 -stim_label 6 dS \
  -stim_file 7 motion.1D'[4]' -stim_base 7 -stim_label 7 dL \
  -stim_file 8 motion.1D'[5]' -stim_base 8 -stim_label 8 dP \
  -gltsym 'SYM: Vrel -Arel' -glt_label 1 V-A \
  -tout -x1D rall_X.xmat.1D -xjpeg rall_X.jpg \
  -fitts rall_fitts -bucket rall_func \
  -jobs 2

```

- 2 audiovisual stimulus classes were given using **-stim\_times**

## • Important to include motion parameters as regressors?

- › May remove the confounding effects due to motion artifacts
- › 6 motion parameters as covariates via **-stim\_file + -stim\_base**
- › **motion.1D** generated from **3dvolreg** with the **-1Dfile** option
- › Test the significance of head motion parameters
  - › Add **-bout** or remove **-stim\_base**
  - › Use **-gltsym 'SYM: roll \ pitch \ yaw \ dS \ dL \ dP'**

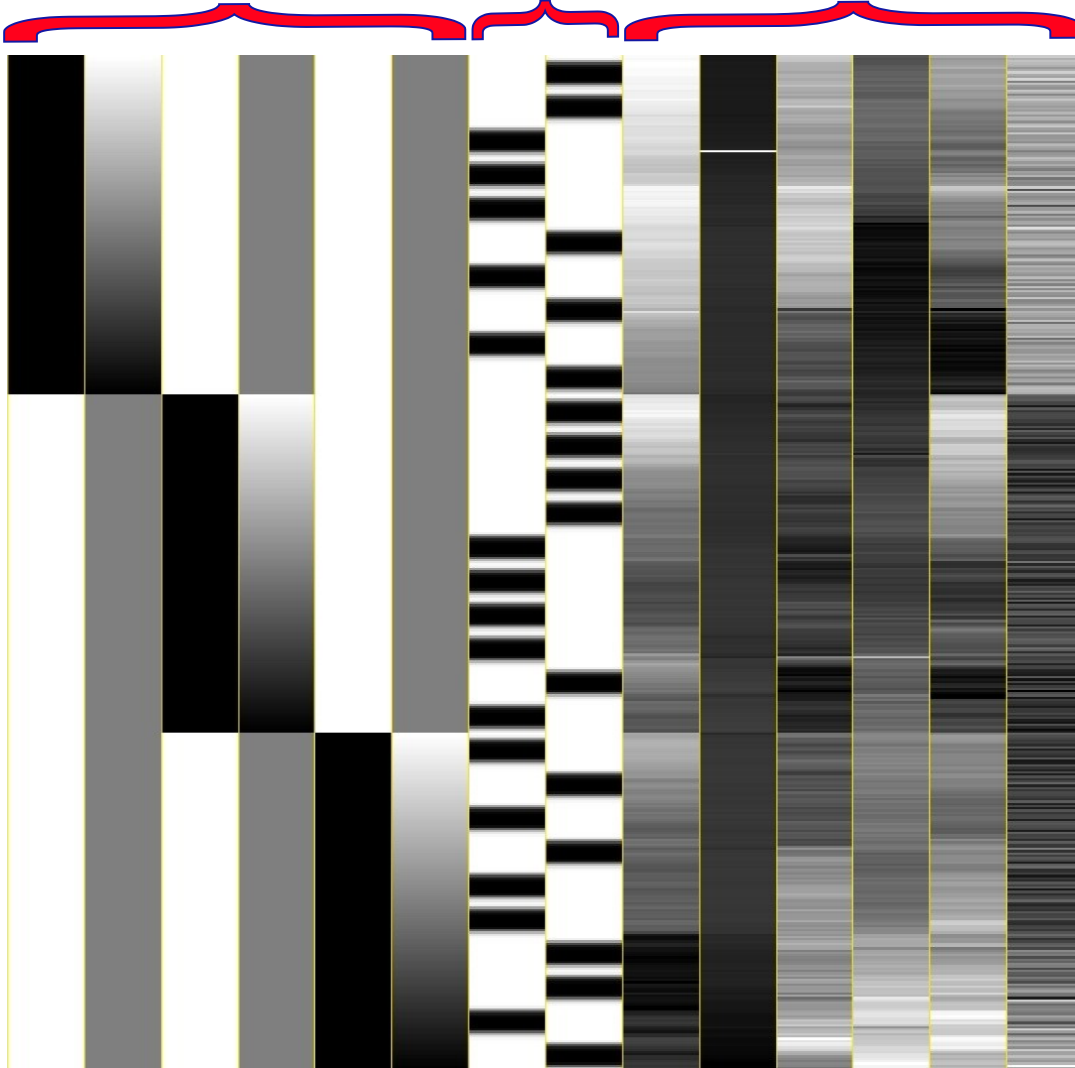
# Modeling Serial Correlation in the Residuals

- Temporal correlation exists in the residuals of the time series regression model
- Within-subject variability (or statistical value) would get deflated (or inflated) if temporal correlation is not accounted for in the model
- Better correct for the temporal correlation if bringing both effect size and within-subject variability to group analysis
- ARMA(1, 1) assumed in **3dREMLfit**
- Script automatically generated by 3dDeconvolve (may use `-x1D_stop`)
  - ★ File **rall\_func.REML\_cmd** under **AFNI\_data6/afni**
  - ▯ Run it by typing **tcsh -x rall\_func.REML\_cmd**

```
3dREMLfit -matrix rall_X.xmat.1D -input rall_vr+orig \  
-tout -Rbuck rall_func_REML -Rvar rall_func_REMLvar \  
-Rfitts rall_fitts_REML -verb
```

# Regressor Matrix X for This Script (via `-xjpeg`)

Baseline    Audiovisual stimuli    Head Motion

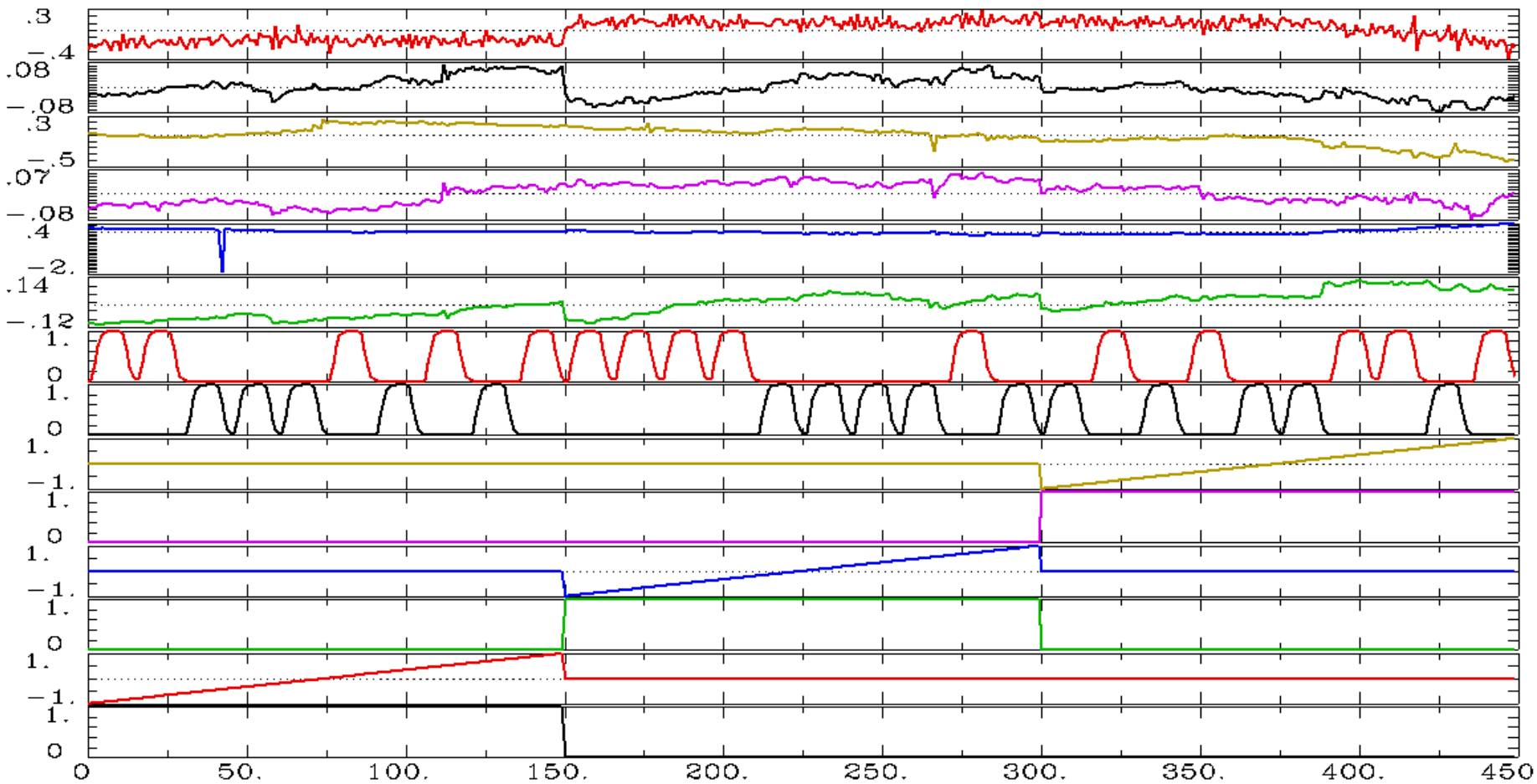


- 6 drift effect regressors
  - linear baseline
  - 3 runs times 2 params/run
- 2 regressors of interest
- 6 head motion regressors
  - 3 rotations and 3 shifts

`aiv_rall_xmat.jpg`

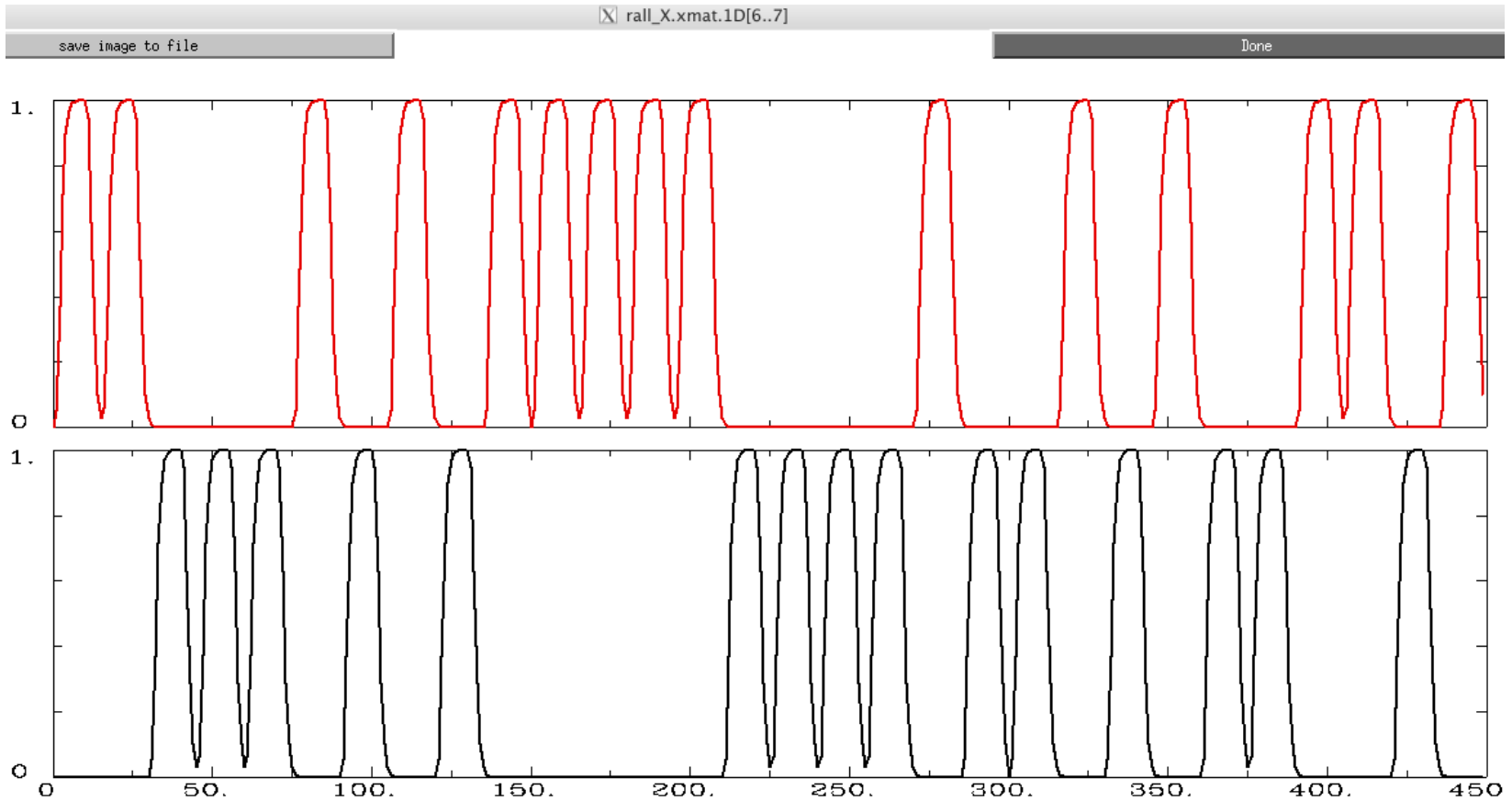
# Showing All Regressors (via -x1D)

save image to file Done



All regressors: **1dplot -sepscl rall\_X.mat.1D**

# Plotting Regressors of Interest



Regressors of Interest: `1dplot rall_X.mat.1D'[6..7]'`

# Options in 3dDeconvolve - 1

**-concat '1D: 0 150 300'**

- “File” that indicates where distinct imaging runs start inside the input file
  - Numbers are the time (TR) **indexes** inside the dataset file for start of runs
  - These time points are considered as **discontinuities** in the model
  - In this case, a text format .1D file put directly on the command line
    - Could also be a filename, if you want to store that data externally

**-num\_stimts 8**

- 2 audiovisual stimuli (+6 motion), thus 2 **-stim\_times** below
- Times given in the **-stim\_times** files are *local* to the start of each run

**-stim\_times 1 stim\_AV1\_vis.txt 'BLOCK(20,1)' -stim\_label 1 Vrel**

- Content of **stim\_AV1\_vis.txt**

```
60 90 120 180 240
120 150 180 210 270
0 60 120 150 240
```

★ Each of 3 lines specifies start time in **seconds** for stimuli within the run

## Options in 3dDeconvolve - 2

```
-gltsym 'SYM: Vrel -Arel' -glt_label 1 V-A
```

- **GLTs**: General Linear Tests
- **3dDeconvolve** provides test statistics for each regressor separately, but to test combinations of the  $\beta$  weights in each voxel, we need **-gltsym** option
- Example above tests the difference between the  $\beta$  weights for the **Virtual-reliable** and the **Audio-reliable** responses
  - **SYM**: means symbolic input is on command line
    - Otherwise inputs will be read from a file
  - Symbolic names for each regressor taken from **-stim\_label** options
  - Stimulus label can be preceded by **+** or **-** to indicate sign to use in combination of  $\beta$  weights
  - **Leave space after each label!**
- Goal is to test a linear combination of the  $\beta$  weights
  - Null hypothesis  $\beta_{Vrel} = \beta_{Arel}$
  - e.g., does **Vrel** get different response from **Arel**?
- What do **'SYM: 0.5\*Vrel +0.5\*Arel'** and **'SYM: Vrel \ Arel'** test?

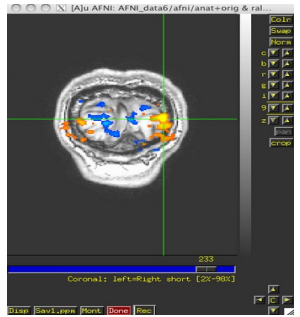
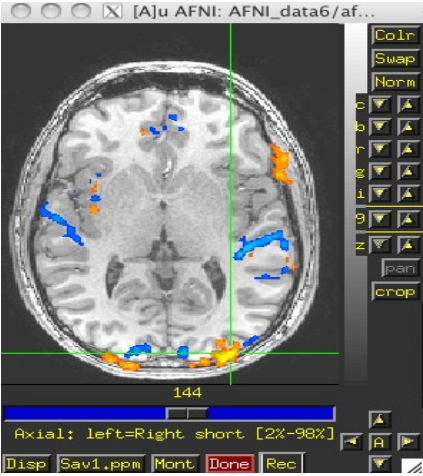
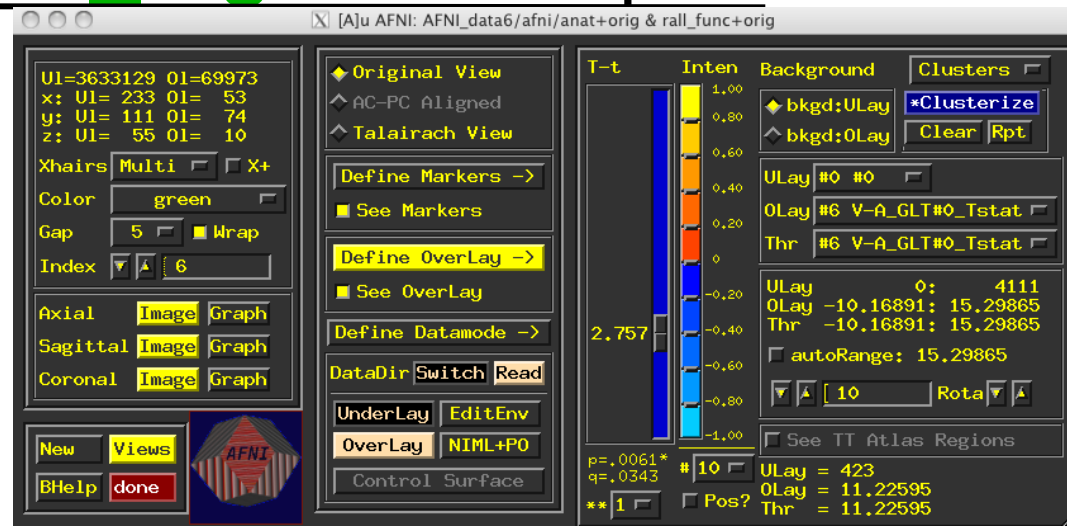
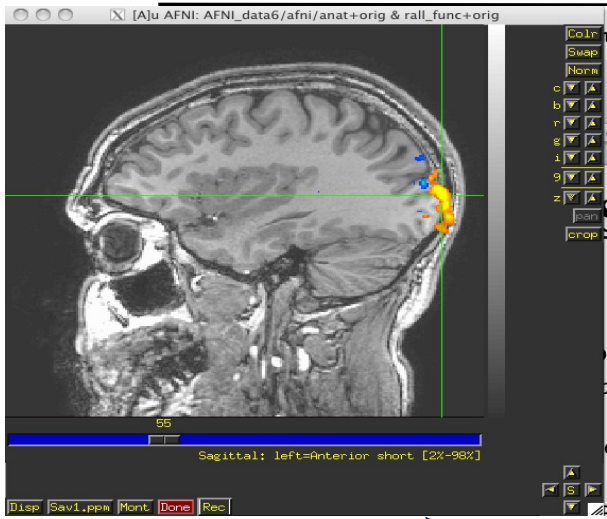
## Options in 3dDeconvolve - 3

**-fout -tout** = output both  $F$ - and  $t$ -statistics for each stimulus class (**-fout**) and stimulus coefficient (**-tout**) — but not for the baseline coefficients (use **-bout** for baseline)

- The full model statistic is an  $F$ -statistic that shows how well all the regressors of interest explain the variability in the voxel time series data
  - Compared to how well *just* the baseline model time series fit the data times (in this example, we have 12 baseline regressor columns in the matrix — 6 for the linear drift, plus 6 for motion regressors)
  - $F = [SSE(r) - SSE(f)] / df(n) \div [SSE(f) / df(d)]$
- The individual stimulus classes also will get individual  $F$ - (if **-fout** added) and/or  $t$ -statistics indicating the significance of their individual *incremental* contributions to the data time series fit
  - If  $DF=1$  (e.g.,  $F$  for a single regressor),  $t$  is equivalent to  $F$ :  $t(n) = F^2(1, n)$



# Results of **rall\_regress** Script



• Images showing results from third GLT contrast: **VrelvsArel**

• Menu showing labels from **3dDeconvolve**

• Play with these results yourself!

# Compare 3dDeconvolve and 3dREMLfit

Terminal — afni — 74x30

```

You have mail.
gangc@fingol:~> cd AFNI_data6/afni
gangc@fingol:~/AFNI_data6/afni> afni &
[1] 40244
gangc@fingol:~/AFNI_data6/afni>
Thanks go to LR Frank for useful feedback

GPL AFNI: Analysis of Functional NeuroImages, by RW Cox (rwcox@nih.gov)
This is Version AFNI_2009_12_31_1431
[[Precompiled binary macosx_10.6_Intel_64: Mar 18 2010]]

** This software was designed to be used only for research purposes. **
** Clinical uses are not recommended, and have never been evaluated. **
** This software comes with no warranties of any kind whatsoever, **
** and may not be useful for anything. Use it at your own risk! **
** If these terms are not acceptable, you aren't allowed to use AFNI.**
** See 'Define Datamode->Misc->License Info' for more details. **

**** If you DO find AFNI useful, please cite this paper:
RW Cox. AFNI: Software for analysis and visualization of
functional magnetic resonance neuroimages.
Computers and Biomedical Research, 29:162-173, 1996.

**** If you find SUMA useful, citing this paper also would be nice:
ZS Saad, RC Reynolds, B Argall, S Japee, RW Cox.
SUMA: An Interface For Surface-Based Intra- And Inter-Subject Analysis
With AFNI. 2nd IEEE International Symposium on Biomedical Imaging:
Macro to Nano 2, 1510-1513, 2004.

Initializing: X11.
  
```

**Group Analysis:** will be carried out on  $\beta$  or GLT coef (+ $t$ -value) from single-subject analysis