

# Deconvolution of Compartmental Water Diffusion Coefficients in Stroke Using DW-IR MEMRI

G. Nair<sup>1,2</sup>, M. Shazeeb<sup>1,2</sup>, J. Bouley<sup>3</sup>, K. G. Helmer<sup>1</sup>, M. Fisher<sup>3,4</sup>, C. H. Sotak<sup>1,4</sup>

<sup>1</sup>Biomedical Engineering, Worcester Polytechnic Institute, Worcester, MA, United States, <sup>2</sup>GSBS, University of Massachusetts Medical School, Worcester, MA, United States, <sup>3</sup>Neurology, UMassMemorial Health Care, Worcester, MA, United States, <sup>4</sup>Radiology, UMassMemorial Health Care, Worcester, MA, United States

**Introduction:** The apparent diffusion coefficient (ADC) of brain water is known to decrease in ischemia.<sup>1</sup> Although the mechanism of the decrease is not well understood, it is thought to be related to relative changes in the intra- and extra-cellular volume fractions during ischemia.<sup>2,3</sup> We sought to identify the relative contributions of intra- and extracellular water ADC changes during ischemia using Mn<sup>2+</sup> as an intracellular MRI contrast agent. Intracerebroventricular (ICV) infusion of Mn<sup>2+</sup> results in neuronal uptake, selectively shortening the T<sub>1</sub> and T<sub>2</sub> relaxation times of intracellular water. The relative differences in T<sub>1</sub> or T<sub>2</sub> relaxation times between the two compartments can be used to selectively null the signal from one of the compartments.<sup>4</sup> In the first set of experiments, the average T<sub>1</sub> and T<sub>2</sub> relaxation times of water were measured in the intra- and extracellular compartments post-Mn<sup>2+</sup> infusion. Secondly, diffusion-weighted inversion-recovery (DW-IR) images were acquired from each compartment by choosing an inversion time (TI) to selectively null the MRI signal from the other compartment.

**Methods:** All MR imaging was performed on a Bruker Biospin 2T/45 cm horizontal bore system equipped with 200mT/m gradients and a surface coil for RF transmit and receive. Two groups of rats (SD, 275-300g) were used in this study. The first group (n=5) was anesthetized with intraperitoneal injection of chloral hydrate (400 mg/kg) and immobilized on a stereotaxic frame. Manganese, 50 µl of 25 mM MnCl<sub>2</sub> in saline, was infused ICV over 2 mins through a burr hole the skull (0.8 mm posterior, 1.2 mm lateral to the bregma, and 4mm below the skull). T<sub>1</sub> was measured using an IR sequence acquired at 2, 12, 24, 48, 72 and 96 hrs time-points post-ICV infusion of Mn<sup>2+</sup>. A sech pulse was used for adiabatic spin-inversion and TI values (21) were spaced logarithmically from 14.88-5000 ms. Other imaging parameters were TR = 7.5 s, TE = 7 ms, FOV = 31 mm, 64 × 64 data matrix with an in-plane resolution of 484 microns and 5 slices of 1mm thickness. T<sub>2</sub> was measured using a CPMG sequence with identical resolution (TR = 5 s, TE varied in 24 steps between 4.5 ms and 108 ms). Bi-exponential T<sub>1</sub> and T<sub>2</sub> maps were calculated using Matlab.

In a second group (n=4), Mn<sup>2+</sup> was infused (ICV) 72 hrs prior to middle cerebral artery occlusion (MCAO). MRI was performed 2 hrs post-MCAO. Eleven DW-IR MRIs were acquired with b-values from 5-1080 s/mm<sup>2</sup> and TR = 5 s, TE = 27 ms, FOV = 31 mm, 64 × 64 matrix acquisition (in-plane resolution of 484 microns), 5 1-mm-thick slices, δ = 8 ms, Δ = 13 ms and diffusion gradients applied in three directions simultaneously. TI values of 100 ms and 450 ms were used to null the intra- and extracellular water signals, respectively. Average ADC was calculated from each brain hemisphere and compared across all 5 slices. A single-tailed, paired-t-test analysis was used; p<0.05 was considered statistically significant.

**Results and Discussion:** T<sub>1</sub>-weighted images acquired at various time points showed uniform enhancement in the brain at 72 hours post-ICV Mn<sup>2+</sup> infusion (the time point used for all experiments). The average bi-exponential T<sub>1</sub> values were T<sub>1a</sub> ~ 150 ms; M<sub>0a</sub> = 64%; T<sub>1b</sub> ~ 700 ms; M<sub>0b</sub> = 34% (**Fig 1**); the bi-exponential T<sub>2</sub> values were T<sub>2a</sub> ~ 9 ms; M<sub>0a</sub> = 66%; T<sub>2b</sub> ~ 51 ms; M<sub>0b</sub> = 34%. Extracellular ADC values calculated from DWIs with TI = 100 ms (intracellular water signal nulled) decreased significantly from 1.3 × 10<sup>-3</sup> mm<sup>2</sup>/s to 1.1 × 10<sup>-3</sup> mm<sup>2</sup>/s during ischemia (**Fig 2**). Similarly, intracellular ADC values calculated from DWIs with TI = 450 ms (extracellular water signal nulled) showed a significant reduction from 0.88 × 10<sup>-3</sup> mm<sup>2</sup>/s to 0.72 × 10<sup>-3</sup> mm<sup>2</sup>/s in the ischemic hemisphere (**Fig 3**). Since Mn<sup>2+</sup> distribution was not uniform in all brain regions, there may be some signal cross-contamination between compartments if each component is not completely nulled at their respective TI values.

**Conclusion:** ICV infusion of Mn<sup>2+</sup> appears to achieve sufficient intracellular concentrations of Mn<sup>2+</sup> to move the inter-compartmental equilibrium water exchange into the slow-exchange regime<sup>5</sup>. When this is the case, it is in principle possible to separate the intra- and extracellular water signals on the basis of differences in their respective T<sub>1</sub> and T<sub>2</sub> relaxation times. The ADCs measured separately from intra- and extracellular water in ischemic rat brain showed a significant reduction during cerebral ischemia. However, the absolute values of both the intra- and extracellular ADCs are somewhat higher than the combined rat-brain ADCs measured from both compartments (~0.8 × 10<sup>-3</sup> mm<sup>2</sup>/s). Furthermore, the percent-ADC reduction following ischemia is not as large in these studies (~15-18%) as that observed from the combined compartments (~30-35%) under similar ischemic conditions. However, the effect of inter-compartmental equilibrium water exchange is not accounted for in either of these cases and this issue may be responsible for some of these differences.

**Reference:** 1) Moseley et al MRM 1990;14:330 2) Neil JJ et al MRM 1996;35:329 3) Duong TQ et al MRM 1998;40:1 4) Silva et al MRM 2002;48:826 5) Labadie et al JMR Ser B 1994;105:99

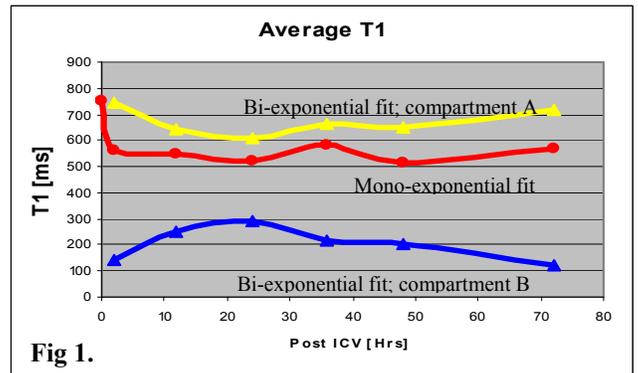


Fig 1.

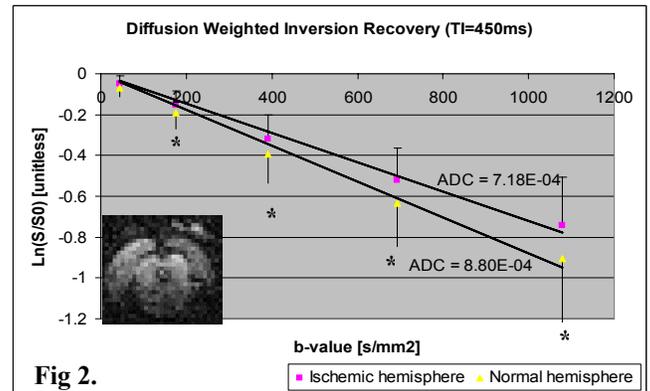


Fig 2.

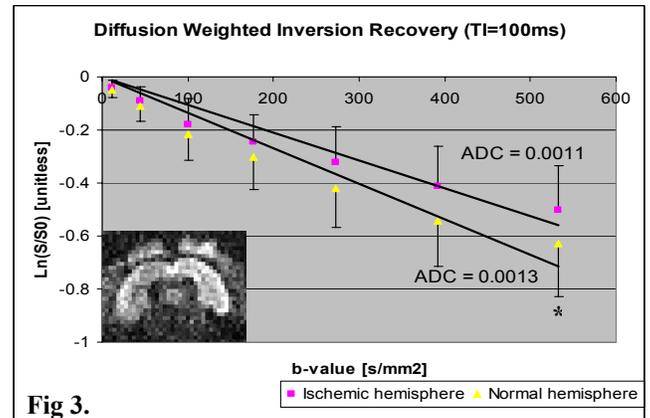


Fig 3.